

Environmental Methicillin-resistant *Staphylococcus aureus* Contamination, Persistent Colonization, and Subsequent Skin and Soft Tissue Infection

Patrick G. Hogan, MPH; Ryan L. Mork, PhD; Ryley M. Thompson; Carol E. Muenks, BA; Mary G. Boyle, MSN; Melanie L. Sullivan, BS; John J. Morelli, BS; Caroline V. Williams, BHS; Nataly Sanchez, BS; David A. Hunstad, MD; Juliane Bubeck Wardenburg, MD, PhD; Sarah J. Gehlert, PhD; Carey-Ann D. Burnham, PhD; Andrey Rzhetsky, PhD; Stephanie A. Fritz, MD, MSCI

[+ Supplemental content](#)

IMPORTANCE The longitudinal association among persistent *Staphylococcus aureus* colonization, household environmental contamination, and recurrent skin and soft tissue infection (SSTI) is largely unexplored to date.

OBJECTIVES To identify factors associated with persistent *S aureus* colonization and recurrent SSTI in households with children with community-associated methicillin-resistant *S aureus* (MRSA) SSTI.

DESIGN, SETTING, AND PARTICIPANTS This 12-month prospective cohort study included 150 children with community-associated MRSA SSTI, 542 household contacts, and 154 pets enrolled from January 3, 2012, through October 20, 2015. A total of 5 quarterly home visits were made to 150 households in the St Louis, Missouri, region. Statistical analysis was performed from September 18, 2018, to January 7, 2020.

EXPOSURES Covariates used in *S aureus* strain persistence and interval SSTI models included *S aureus* colonization and contamination measures, personal hygiene and sharing habits, health history, activities external to the home, and household characteristics (eg, cleanliness, crowding, home ownership, and pets). Serial samples to detect *S aureus* were collected from household members at 3 anatomic sites, from pets at 2 anatomic sites, and from environmental surfaces at 21 sites.

MAIN OUTCOMES AND MEASURES Molecular epidemiologic findings of *S aureus* isolates were assessed via repetitive-sequence polymerase chain reaction. Individual persistent colonization was defined as colonization by an identical strain for 2 consecutive samplings. Longitudinal, multivariable generalized mixed-effects logistic regression models were used to assess factors associated with persistent *S aureus* personal colonization, environmental contamination, and interval SSTI.

RESULTS Among 692 household members in 150 households, 326 (47%) were male and 366 (53%) were female, with a median age of 14.82 years (range, 0.05-82.25 years). Of 540 participants completing all 5 samplings, 213 (39%) were persistently colonized with *S aureus*, most often in the nares and with the strain infecting the index patient at enrollment. Nine pets (8%) were persistently colonized with *S aureus*. Participants reporting interval intranasal mupirocin application were less likely to experience persistent colonization (odds ratio [OR], 0.44; 95% credible interval [CrI], 0.30-0.66), whereas increasing strain-specific environmental contamination pressure was associated with increased individual persistent colonization (OR, 1.17; 95% CrI, 1.06-1.30). Strains with higher colonization pressure (OR, 1.47; 95% CrI, 1.25-1.71) and MRSA strains (OR, 1.57; 95% CrI, 1.16-2.19) were more likely to persist. Seventy-six index patients (53%) and 101 household contacts (19%) reported interval SSTIs. Individuals persistently colonized with MRSA (OR, 1.56; 95% CrI, 1.17-2.11), those with a history of SSTI (OR, 2.55; 95% CrI, 1.88-3.47), and index patients (OR, 1.54; 95% CrI, 1.07-2.23) were more likely to report an interval SSTI.

CONCLUSIONS AND RELEVANCE The study findings suggest that recurrent SSTI is associated with persistent MRSA colonization of household members and contamination of environmental surfaces. Future studies may elucidate the effectiveness of specific combinations of personal decolonization and environmental decontamination efforts in eradicating persistent strains and mitigating recurrent SSTIs.

JAMA Pediatr. doi:10.1001/jamapediatrics.2020.0132
Published online March 30, 2020.

Author Affiliations: Author affiliations are listed at the end of this article.

Corresponding Author: Stephanie A. Fritz, MD, MSCI, Department of Pediatrics, Washington University School of Medicine in St Louis, 660 S Euclid Ave, Campus Box 8116, St Louis, MO 63110 (fritz.s@wustl.edu).

S *taphylococcus aureus* is the most common cause of skin and soft tissue infection (SSTI) in the United States.^{1,2} The emergence of community-associated methicillin-resistant *S aureus* (MRSA) in recent decades has posed unique infection prevention challenges because community-associated MRSA establishes community reservoirs and spreads via asymptomatic colonization.^{3,4}

States of *S aureus* colonization include noncarriage, intermittent colonization, and persistent colonization.^{5,6} Because colonization is associated with increased risk for infection,⁷ understanding these states is central to addressing the epidemic of community-associated MRSA infection. Although colonization can resolve spontaneously or with topical antimicrobial use,^{5,6} the reported median duration of MRSA colonization is approximately 4 months despite systemic antibiotic and topical antimicrobial use.⁸ Persistently colonized individuals can be sources of transmission to others and their environment and thus warrant specific attention.

Studies⁹⁻¹¹ have shown that community-associated MRSA infection is frequently spread among households, with personal *S aureus* colonization, environmental contamination, and pet carriage as reservoirs.¹²⁻¹⁶ Recurrent SSTIs in index patients and colonization and subsequent infection in household contacts are common and concerning.^{9,10,14,17,18} Recent studies^{9,13,17} showed an SSTI recurrence rate of 43% to 51% in index patients within 6 months and subsequent SSTI in 13% of household contacts. Baseline MRSA environmental contamination was a risk factor for recurrent SSTIs in index patients^{13,17} and likewise was associated with recurrent environmental contamination¹⁹ and human reacquisition²⁰ in the 3 months after baseline infection.

Although previous studies^{9,13,17,19,20} provided insights into persistent colonization and recurrent infection, to date, little is known about their association over time. We investigated factors associated with persistent *S aureus* colonization and recurrent SSTI for 1 year among households with children with community-associated MRSA SSTI to ascertain the association among environmental contamination, persistent personal colonization, and recurrent SSTI and to identify targets for intervention to disrupt the household reservoir that perpetuates colonization and recurrent SSTIs. We thus used household-level, high-resolution molecular typing with comprehensive, longitudinal sampling of household members, environmental surfaces, and pets.

Methods

Participants and Data Collection

Patients with culture-confirmed, community-associated MRSA SSTI were recruited for the Household Observation of MRSA in the Environment (HOME) study^{12,15,16,21} from hospitals and community practices in metropolitan St Louis, Missouri, from January 3, 2012, through October 20, 2015. We conducted a baseline visit at the index patient's primary home at a median of 20 days (interquartile range [IQR], 13-29 days) after infection. Household contacts who slept in the home at least 4 nights per week and dogs and cats who lived indoors were enrolled.

Key Points

Question Is there an association between environmental contamination, persistent personal colonization, and recurrent skin and soft tissue infection among households with children with community-associated methicillin-resistant *Staphylococcus aureus* (MRSA) skin and soft tissue infection?

Findings In this cohort study of 692 household members from 150 households with children with community-associated MRSA skin and soft tissue infection, 39% of participants were persistently colonized with *S aureus*. A total of 53% of index patients and 19% of household contacts reported interval skin and soft tissue infections, more often when persistently colonized with MRSA.

Meaning The findings suggest that MRSA colonization of household members and contamination of environmental surfaces in the household may be associated with MRSA skin and soft tissue infections in children.

Written informed consent and assent were obtained for household members and pets; study procedures were approved by the Washington University institutional review board and Institutional Animal Care and Use Committee, St Louis, Missouri. Data were analyzed from September 18, 2018, to January 7, 2020.

Baseline surveys were administered to each household member to assess *S aureus* infection history, hygiene practices, activities outside the home, pet and household characteristics, and household cleaning habits. A household cleanliness score, adapted from The Environmental Cleanliness and Clutter Scale,²² (score: 1 [above average], 2 [average], 3 [below average], and 4 [very dirty]), was assigned by the research team to control potential reporting bias about cleaning practices. At 4 quarterly follow-up visits, participants were asked about interval SSTIs, emergency department and hospital visits, and systemic and topical antimicrobial use. Medical records were requested for treated SSTIs.

Samples were obtained from the anterior nares, axillae, and inguinal folds of household members (Eswab; Becton Dickinson), nares (minutip Eswab; Becton Dickinson) and dorsal fur (Eswab) of indoor pets as well as up to 21 environmental surfaces to test for *S aureus* at each visit. Sampled environmental surfaces (Eswabs and Baird Parker Agar contact plates [Hardy Diagnostics]) included shared electronics (n = 4) and kitchen (n = 5), bathroom (n = 11), and bedroom (n = 1) surfaces (eTable 1 in the [Supplement](#)).²³

Microbiological Methods

Eswabs were processed using broth-enrichment (Becton Dickinson) culture methods; from contact plates, colonies were selected based on morphologic features. *S aureus* was isolated and antibiotic susceptibility testing was performed using established techniques (eMethods in the [Supplement](#)).²⁴ When available, SSTI isolates from the index patients' enrollment infections (infecting strains; n = 91) and interval SSTI isolates from household members (n = 19) were obtained from clinical microbiologic laboratories; alternatively, antibiotic

susceptibility profiles were obtained. To discern distinct *S aureus* strains, molecular typing was performed via repetitive-sequence polymerase chain reaction using a 95% similarity cutoff.^{25,26}

Definitions

Strains were defined based on the composite of repetitive-sequence polymerase chain reaction designation and methicillin-resistance profile for each *S aureus* isolate recovered within the context of a single household. An individual persistent colonization event was colonization of an individual by an identical strain for 2 consecutive samples from 1 or more anatomic sites. Colonization with an acquired novel strain did not constitute persistent colonization (ie, persistence). Persistence degree was considered to be the number of consecutive persistence events observed (ie, number of consecutive colonization events minus 1). Colonization at 2, 3, 4, or 5 consecutive samplings represented a persistence degree of 1, 2, 3, or 4, respectively.

Normalized persistence events were the number of persistence events divided by the number of potential persistence events (maximum of 4 events) \times 100. Events were not necessarily consecutive in contrast to persistence degree.

In personal persistence, a strain was observed colonizing 1 or more individuals at 2 or more consecutive samplings. In environmental persistence, a strain was found contaminating any set of household environmental sites at consecutive samplings. In household persistence, a strain persisted across any personal, pet, and/or environmental sites at consecutive samplings.

Personal colonization pressure was the number of anatomic sites (3 per person) colonized with *S aureus*, methicillin-susceptible *S aureus* (MSSA), MRSA, or a given strain divided by the total number of anatomic sites sampled per household. Environmental contamination pressure was the number of contaminated surfaces divided by the total number of surfaces sampled per household.

Statistical Analysis

Univariate analyses were conducted in Python 2.7.15 (Python Software Foundation) using the *scipy* package 1.1.0²⁷ or in R (R Foundation for Statistical Computing).²⁸ Categorical data were analyzed with Fisher exact test. For continuous data, Kruskal-Wallis nonparametric 1-way analysis of variance was used for pairwise comparisons and Spearman's ρ was calculated to determine correlations. All statistical tests were 2-sided.

The individual persistence, interval SSTI, household persistence, and infecting-strain persistence models were bayesian, longitudinal, multivariable generalized mixed-effects logistic regression models fitted using R library *MCMCglmm*.²⁹ Random intercepts were included to control for repeated longitudinal sampling at the individual, household, and strain-type levels. The threshold family link function was used with a χ^2 prior for random intercepts and a normal prior for fixed effects. A heuristic, 2-stage covariate selection scheme driven by domain expertise controlled for the large number of covariates measured. In the first stage, 11 covariates were selected

based on a priori hypotheses and statistically significant variables in univariate analysis for each outcome. All possible combinations were fitted and the optimal model selected based on the lowest deviance information criterion. In the second stage, each model was reestimated with the individual addition of other covariates of interest, which were maintained in the final, presented models when model deviance information criterion was reduced and the covariate exhibited a Markov chain Monte Carlo $P < .05$. eTables 2-5 in the [Supplement](#) list primary and secondary model covariates; the eMethods in the [Supplement](#) gives further details.

A first-order Markov model of household strain persistence was fitted using R library *msm*³⁰ (eMethods in the [Supplement](#)). This model included 5 states to describe colonization by a given strain within a household: at least 1 child, at least 1 adult, or both a child and adult colonized, exclusively environmental contamination, and no colonization or contamination. Transition probabilities and 95% CI estimates are presented for comparison.

Results

Study Population and Longitudinal Colonization Status

From January 3, 2012, through October 20, 2015, 150 households were enrolled in a 12-month longitudinal study, including 150 index patients (70 [47%] male; median age, 2.99 years [range, 0.08-18.57 years]) presenting with community-associated MRSA infection, their 542 household contacts (256 [47%] male; median age, 25.80 years [range, 0.05-82.25 years]), and their 154 indoor pets (116 dogs, 38 cats) (eTable 6 in the [Supplement](#)). Median household size was 4 persons (range, 2-13 persons). Of 150 households enrolled, 135 (90%) completed the 12-month study visit; eTable 6 in the [Supplement](#) provides further details of longitudinal sampling completion for household members, environmental surfaces, and pets. A total of 513 of 692 participants (74%) were colonized with *S aureus* at least once, 319 (46%) were colonized with MRSA. Of 150 household environments, 136 (91%) were contaminated with *S aureus* at least once, and 104 (69%) were contaminated with MRSA. Of 154 pets, 68 (44%) carried *S aureus* at least once, and 44 (29%) carried MRSA. Over the study period, 290 of 671 participants (43%) received decolonization treatment at least once: 148 (22%) applied mupirocin to their anterior nares and 243 (36%) performed antiseptic body washes.

Individual *S aureus* Strain Persistence

Of participants completing all 5 samplings ($n = 540$), 306 (57%) were colonized at 2 or more consecutive samples with any strain of *S aureus*; of these, 213 (70%; 213 of 540 [39%] overall) exhibited strain persistence (Table 1). Of 142 individuals (26%) colonized consecutively with any strain of MRSA, 109 (77%) exhibited strain persistence (Figure 1). Of 1171 *S aureus* colonization events with a subsequent sampling, persistent colonization was observed in 471 events (40%). Individuals colonized in the nares were most likely to exhibit persistent colonization (at any site) (eTable 7 in the [Supplement](#)). Persistence degree (number of consecutive persistence events) was

Table 1. Colonization Events and Association With Interval SSTIs

Colonization prevalence and frequency ^a	All individuals (n = 540)		Index patients (n = 128)		Household contacts (n = 412)	
	No. (%) ^b	Positive samplings, median (IQR)	No. (%) ^b	Positive samplings, median (IQR)	No. (%) ^b	Positive samplings, median (IQR)
Any <i>S aureus</i>	420 (78)	2 (1-4)	100 (78)	2 (1-4)	320 (78)	2 (1-4)
MRSA ^c	265 (49)	0 (0-2)	76 (59)	1 (0-2)	189 (46)	0 (0-1)
MSSA	286 (53)	1 (0-2)	65 (51)	1 (0-2)	221 (54)	1 (0-2)
Infecting strain ^d	123 (23)	0 (0-1)	40 (31)	1 (0-1)	83 (20)	0 (0-1)
Strain persistence and degree	No. (%) ^e	Persistence degree, median (IQR) ^f	No. (%) ^e	Persistence degree, median (IQR) ^f	No. (%) ^e	Persistence degree, median (IQR) ^f
Any <i>S aureus</i>	213 (39)	2 (1-3)	52 (41)	1.5 (1-2)	161 (39)	2 (1-3)
MRSA ^c	109 (20)	1 (1-3)	33 (26)	1 (1-3)	76 (18)	1 (1-3)
MSSA	112 (21)	2 (1-2)	22 (17)	1 (1-2)	90 (22)	2 (1-2)
Infecting strain ^d	48 (9)	2 (1-3)	19 (15)	1 (1-3)	29 (7)	2 (1-3)
Association with No. of interval SSTIs ^g	p	P value	p	P value	p	P value
Colonization frequency						
Any <i>S aureus</i>	0.11	.03	0.18	.07	0.09	.10
MRSA ^c	0.29	<.001	0.42	<.001	0.21	<.001
MSSA	-0.14	.005	-0.19	.06	-0.11	.06
Infecting strain ^h	0.36	<.001	0.42	<.001	0.29	<.001
Strain persistence degree						
Any <i>S aureus</i>	0.09	.07	0.18	.08	0.07	.18
MRSA ^c	0.27	<.001	0.35	<.001	0.24	<.001
MSSA	-0.14	.003	-0.18	.07	-0.12	.03
Infecting strain ^h	0.34	<.001	0.40	.001	0.28	<.001

Abbreviations: IQR, interquartile range; MRSA, methicillin-resistant *Staphylococcus aureus*; MSSA, methicillin-susceptible *S aureus*; SSTI, skin and soft tissue infection.

^a Eligible individuals for this analysis were those who underwent all 5 samplings (n = 540).

^b The number of individuals colonized at least once.

^c All MRSA, including infecting-strain MRSA.

^d The isolate recovered from the index patient's enrollment infection (infecting strain) was available in 91 households; thus, the number for the all individuals (n = 312), index patients (n = 77), and household contacts (n = 235) columns

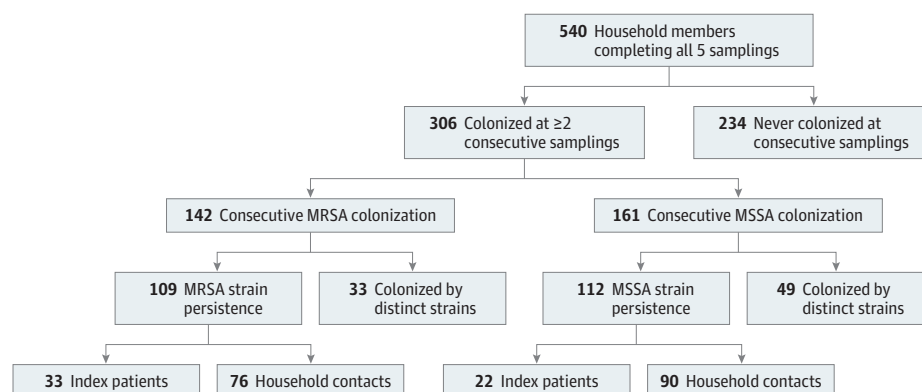
was reduced in rows associated with the infecting strain.

^e The number of individuals colonized with the same strain at consecutive samplings at least once (persistence degree ≥ 1).

^f The median (IQR) is for individuals with persistence degree of 1 or greater.

^g Eligible individuals for this analysis were those who underwent all 5 samplings and were colonized at least once (420 overall, 100 index patients, and 320 household contacts). Spearman correlation test was used.

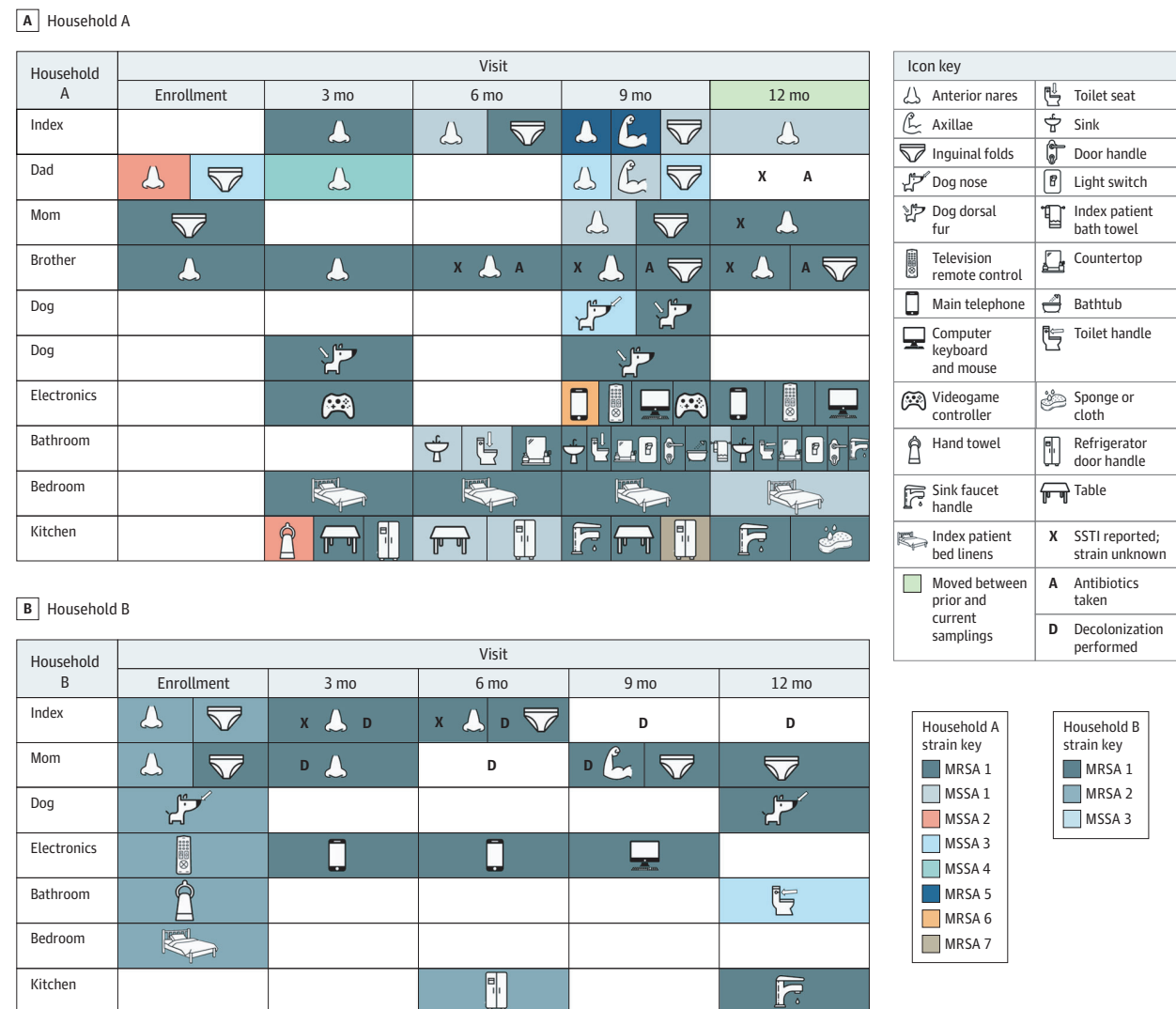
^h The infecting strain was available in 91 households; thus, the number for all individuals (n = 238), index patients (n = 63), and household contacts (n = 175) was reduced in rows associated with the infecting strain.

Figure 1. Flow Diagram of Individual *Staphylococcus aureus* Strain Persistence

Flow diagram of 540 household members completing all 5 samplings; 213 (39%) were persistently colonized with *S aureus* (ie, colonized by an identical *S aureus* strain for ≥ 2 consecutive samplings). Consecutive colonization with methicillin-resistant *S aureus* (MRSA) and then methicillin-susceptible *S aureus* (MSSA) or MSSA and then MRSA occurred; these people were colonized at 2 or more consecutive samplings (n = 306) but did not represent consecutive MRSA

(n = 142) or MSSA (n = 161) colonization. Of note, 8 participants were persistently colonized with both MRSA and MSSA strains throughout the study, and are included in both sides of the flow diagram. Index patients (n = 128) were not more likely than household contacts (n = 412) to be persistently colonized with MRSA or MSSA.

Figure 2. Longitudinal Strain Persistence and Interval Skin and Soft Tissue Infection (SSTI) in Exemplar Households



In households A and B, the infecting strain in the index patient at enrollment was MRSA 1 [methicillin-resistant *S aureus*]. In household B, the index patient had an SSTI with MRSA 1 nineteen months before their enrollment infection; the

infection reported at their 6-month follow-up visit was also MRSA 1. Decolonization measures included nasal mupirocin, bleach baths, or chlorhexidine body washes. MSSA indicates methicillin-susceptible *S aureus*.

similar for MRSA (median, 2 events [IQR, 2-4 events]) and MSSA (3 events [IQR, 2-4 events]; $P = .80$) (eFigure 1 in the [Supplement](#)). Environmental surfaces infrequently exhibited persistence compared with people (1.9% vs 11.7% normalized persistence events; $P < .001$) (eFigure 2 and eTable 1 in the [Supplement](#)). Among 91 households in which the infecting MRSA strain was available, index patients were more likely to be persistently colonized with the infecting strain than were household contacts (7.6% vs 3.8% normalized persistence events; $P = .02$) (eFigure 2 in the [Supplement](#)). Exemplar longitudinal strain persistence is illustrated in Figure 2 and eFigure 3 in the [Supplement](#).

In univariate analysis, eczema diagnosis (odds ratio [OR], 1.77; 95% CI, 1.07-2.94), colonization with the infecting strain at previous sampling (OR, 1.46; 95% CI, 1.04-2.05), and shar-

ing a bath towel (OR, 1.81; 95% CI, 1.21-2.71) were associated with persistence (eTable 8 in the [Supplement](#)). Although interval application of intranasal mupirocin was associated with reduced persistence (OR, 0.39; 95% CI, 0.24-0.65), bleach baths or chlorhexidine body washes were not (OR, 0.96; 95% CI, 0.68-1.36). Fewer persistence events were observed in households that were owned (median, 0.5 [IQR, 0.2-1.0]) vs rented (median, 1.0 [IQR, 0.5-1.7]; $P < .001$) or were rated clean per the interviewers' household cleanliness score (0.6 [IQR, 0.2-1.2] vs 0.9 [IQR, 0.5-1.5]; $P = .03$). The number of persistence events correlated with the number of individuals per square foot in a household (ρ , 0.24; $P = .004$), warmer weather (ρ , 0.11; $P = .02$), and personal *S aureus* colonization pressure (ρ , 0.43; $P < .001$) and environmental *S aureus* contamination pressure (ρ , 0.27; $P < .001$) at previous sampling.

Table 2. Factors Retained in Persistence and Interval SSTI Multivariable Models^a

Model, covariate	Odds ratio (95% credible interval)	Markov chain Monte Carlo P value
Individual persistence^b		
Interval application of intranasal mupirocin vs no mupirocin application	0.44 (0.30-0.66)	<.001
Environmental strain contamination pressure (increase of 1 SD, 0.16, from the mean 0.11)	1.17 (1.06-1.30)	<.001
Frequent handwashing score vs infrequent ^c	1.39 (1.11-1.74)	.003
Personal colonization pressure of other household strains (increase of 1 SD, 0.15, from the mean 0.12)	0.86 (0.77-0.96)	.002
Home ownership vs renting	0.75 (0.59-0.97)	.03
Towel washed after each use vs less frequently	0.81 (0.65-1.01)	.06
Shares brush or comb vs does not share	0.85 (0.68-1.06)	.17
Infecting strain persistence^d		
Interval application of intranasal mupirocin, proportion of household members (increase of 1 SD, 0.28, from the mean 0.09)	0.46 (0.27-0.78)	.002
Interval bleach baths, proportion of household members (increase of 1 SD, 0.29, from the mean 0.14)	2.09 (1.15-3.80)	.005
Interval SSTI, proportion of household members (increase of 1 SD, 0.21, from the mean 0.15)	1.60 (1.10-2.33)	.01
People per bedroom (increase of 1 SD, 0.57, from the mean 1.46)	0.62 (0.34-1.11)	.08
Home ownership vs renting	0.62 (0.18-1.80)	.40
Interval chlorhexidine body washes, proportion of household members (increase of 1 SD, 0.19, from the mean 0.05)	0.89 (0.55-1.38)	.60
Household persistence^e		
Personal strain colonization pressure (increase of 1 SD, 0.11, from the mean 0.11)	1.47 (1.25-1.71)	<.001
Strain types, No. (increase of 1 SD, 1.33, from the mean 2.27)	1.88 (1.59-2.26)	<.001
Personal colonization pressure of other household strains (increase of 1 SD, 0.15, from the mean 0.15)	0.80 (0.69-0.93)	.003
MRSA strain type vs MSSA	1.57 (1.16-2.19)	.005
Interval systemic antibiotic use, proportion of household members (increase of 1 SD, 0.13, from the mean 0.06)	0.91 (0.78-1.04)	.17
People per 1000 ft ² (increase of 1 SD, 1.89, from the mean 3.67)	0.96 (0.81-1.16)	.63
Home ownership vs renting	0.94 (0.64-1.38)	.76
Interval SSTI^f		
SSTI in past year vs no SSTI in past year	2.55 (1.88-3.47)	<.001
MRSA persistence vs no MRSA persistence	1.56 (1.17-2.11)	.004
MSSA persistence vs no MSSA persistence	0.53 (0.32-0.85)	.007
Index patient vs household contact	1.54 (1.07-2.23)	.02
Environmental MRSA contamination pressure (increase of 1 SD, 0.14, from the mean 0.08)	1.09 (0.99-1.21)	.07
Shares bedroom with MRSA-colonized individual vs does not share	1.23 (0.94-1.60)	.12
Frequent handwashing score vs infrequent ^c	1.07 (0.83-1.42)	.60
Child vs adult	0.92 (0.67-1.25)	.60

Abbreviations: MRSA, methicillin-resistant *Staphylococcus aureus*; MSSA, methicillin-susceptible *S aureus*; SSTI, skin and soft tissue infection.

^a For all observations across models, all values are complete observations with no missing values. Odds ratios for continuous covariates are associated with an increase of 1 SD from the mean (zero-centered).

^b Eligible individuals were those colonized with *S aureus* at 1 sampling and sampled subsequently, totaling 907 observations of 443 individuals in 133 households across samplings.

^c Aggregate variable defined as washing hands always after using bathroom, always before preparing food, at least frequently before eating, and at least frequently after changing a diaper (when applicable).

^d Eligible households were those with an available infecting strain (n = 91) present within the household in at least 1 subsequent sampling, totaling 169 observations in 71 households across samplings.

^e Eligible households included household members and environmental surfaces sampled consecutively at least once with at least 1 *S aureus* strain type present at the first sampling, totaling 780 observations in 119 households across samplings.

^f Eligible individuals were those sampled consecutively at least once, along with sampling of environmental surfaces, totaling 1952 observations of 640 individuals in 142 households across samplings.

In the longitudinal, multivariable generalized mixed-effects logistic regression model, individuals reporting interval application of intranasal mupirocin were less likely to experience persistence (OR, 0.44; 95% credible interval [CrI], 0.30-0.66); those performing frequent handwashing were more likely to exhibit persistence (OR, 1.39; 95% CrI, 1.11-1.74) (Table 2). Increasing environmental strain contamination pressure was associated with increased individual persistence (OR, 1.17; 95% CrI, 1.06-1.30), whereas home ownership (OR, 0.75; 95% CrI, 0.59-0.97) and increasing personal colonization pressure of other strains colonizing household contacts

(OR, 0.86; 95% CrI, 0.77-0.96) were associated with decreased likelihood of persistence (Figure 2A).

Pet Strain Persistence

Of 106 pets (83 dogs, 23 cats) from 57 households with at least 2 consecutive samplings, 9 dogs were persistently colonized with *S aureus*, totaling 13 persistence events. In 7 (54%) of these events, the primary caretaker or household member who shared a bed with the pet was colonized with the persistent strain at the previous sampling (eFigure 3C in the Supplement). Pet age, health status, history of SSTI, and overall

environmental and personal *S aureus* colonization pressure were not significantly associated with persistence in pets (eTable 9 in the [Supplement](#)).

Household Strain Persistence

Across 98 households in which household members and environmental surfaces were sampled at all 5 visits, 69 (70%) had at least 1 *S aureus* strain with a persistence degree of at least 2 (eTable 10 in the [Supplement](#)). Forty-five households (46%) exhibited MRSA persistence degree of at least 2; of these households, the persistent MRSA strain colonized at least 3 individuals at 1 or more sampling in 26 (58%) and contaminated the environment in 44 (98%). The MRSA strain infecting the index patient persisted longer than MSSA or noninfecting MRSA strains (persistence degree: median, 2 [IQR, 0-4] vs 1 [IQR, 0-2]; $P = .01$) (Figure 2 and eTable 10 and eFigure 3 in the [Supplement](#)). Moving to a new dwelling was associated with reduced persistence across household members, pets, and the environment (OR, 0.51; 95% CI, 0.25-0.99) (eFigure 3 in the [Supplement](#)). Neither the frequency of participant-reported environmental surface cleaning nor the assigned household cleanliness score was significantly associated with persistence on specified surfaces. More household members per bathroom was associated with increasing number of persistence events on bathroom surfaces (ρ , 0.22; $P = .02$).

We then modeled the association between persistence of the infecting strain (when known; $n = 91$) and reported interval SSTIs, controlling for socioeconomic status, crowding, and decolonization measures. The proportion of household members reporting interval SSTIs was associated with increased persistence of the index patient's enrollment infecting strain at the household level (OR, 1.60; 95% CrI, 1.10-2.33) (Table 2). The proportion of household members reporting interval application of intranasal mupirocin was associated with reduced persistence of the index patient's enrollment infecting strain at the household level (OR, 0.46; 95% CrI, 0.27-0.78); the proportion of household members reporting interval bleach baths was associated with increased persistence of the index patient's infecting strain at the household level (OR, 2.09; 95% CrI, 1.15-3.80).

In the multivariable household persistence model, MRSA (vs MSSA; OR, 1.57; 95% CrI, 1.16-2.19), strains with higher personal colonization pressure (OR, 1.47; 95% CrI, 1.25-1.71), and strains in households with a higher number of distinct strains (OR, 1.88; 95% CrI, 1.59-2.26) exhibited a higher likelihood of persistence (Table 2). Strains in households with higher personal colonization pressure of other strains (OR, 0.80; 95% CrI, 0.69-0.93) (Figure 2A) were less likely to persist.

Markov Model of Household Strain Persistence Dynamics

A first-order Markov model was constructed to capture the persistence dynamics of *S aureus* strains in households among 5 possible colonization states for adults, children, and the environment (Figure 3 and eMethods in the [Supplement](#)). Strains in both the adult and child states tended to persist, suggesting that strains most commonly remain within households by persistent colonization of 1 individual or by transitions between adults or between children. Strains in the environment-only state showed lower persistence.

Interval SSTI

Of 144 index patients with at least 1 follow-up visit, 76 (53%) reported 164 recurrent SSTIs, with 45 (31%) reporting 2 or more subsequent SSTIs (range, 1-9 SSTIs) (eTable 11 in the [Supplement](#)). Of 523 household contacts, 101 (19%) experienced 169 SSTIs (range, 1-13 SSTIs). Participants sought medical care for 178 of 333 reported SSTIs (53%), for which 80% were prescribed a systemic antibiotic; 19 (7%) interval infecting isolates were available for molecular characterization (eTable 12 in the [Supplement](#)). In 14 households with an available infecting strain at enrollment and interval SSTI strains, 11 isolate pairs (79%) were identical strains. Interval infecting strains were commonly found in the household at the previous sampling (14 of 19 [74%]), often colonizing the individual reporting the SSTI (8 of 19 [42%]) and/or contaminating the environment (9 of 17 [53%]) (Figure 2B and eFigure 3 in the [Supplement](#)). Additional exemplar interval SSTIs are illustrated in Figure 2 and eFigure 3 in the [Supplement](#).

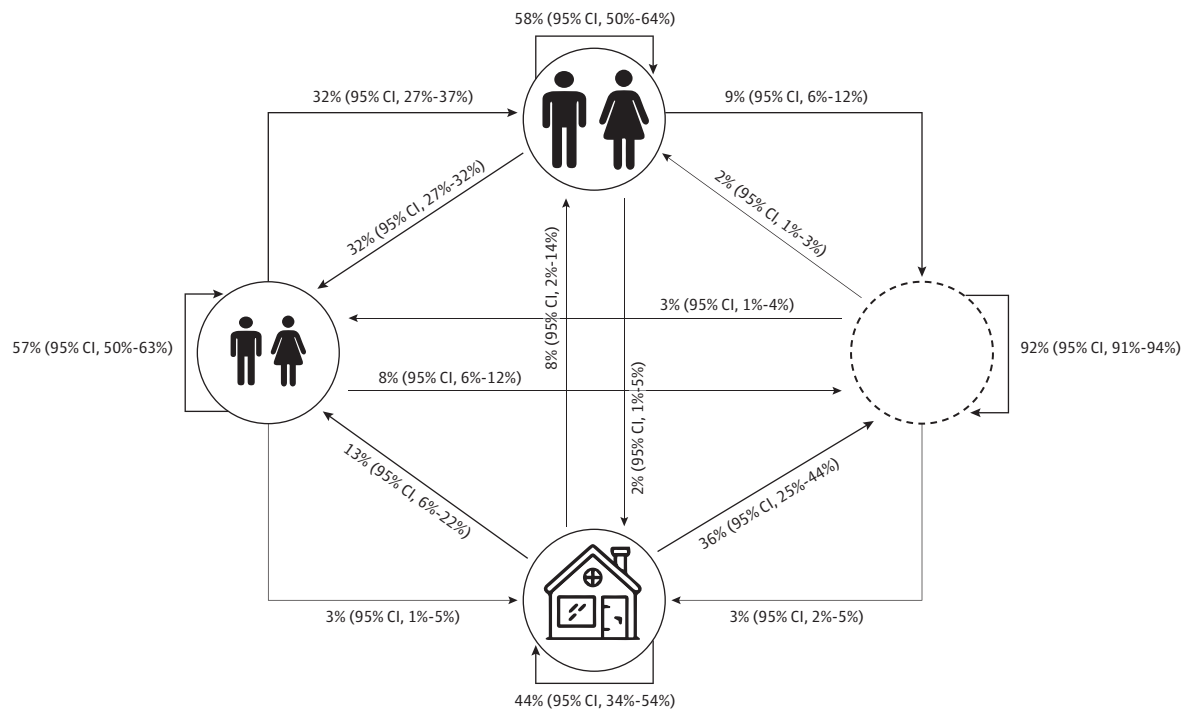
Individuals exhibiting a higher degree of MRSA strain persistence reported more interval SSTIs (ρ , 0.27; $P < .001$) (Table 1 and Figure 2A). Increasing persistence degree of the most persistently MRSA-colonized individual was associated with an increase in the total number of interval household SSTIs (ρ , 0.30; $P < .001$) (eFigure 4 in the [Supplement](#)). A higher degree of MSSA strain persistence was associated with fewer individual SSTIs (ρ , -0.14; $P < .003$) (Table 1 and eFigure 4 in the [Supplement](#)).

In univariate analysis, individuals reporting a history of SSTI in the year before enrollment (OR, 5.79; 95% CI, 3.93-8.53) and children (OR, 1.60; 95% CI, 1.12-2.27) were more likely to report at least 1 interval SSTI during the study (eTable 13 in the [Supplement](#)). Hygiene measures, such as showering (vs bathing; OR, 0.58; 95% CI, 0.40-0.85), frequent handwashing (OR, 0.69; 95% CI, 0.47-0.99), and frequent tooth brushing (OR, 0.59; 95% CI, 0.37-0.94), were inversely associated with interval SSTI. Individuals colonized with MRSA (OR, 2.91; 95% CI, 2.22-3.82) and/or colonized with the infecting strain (OR, 3.55; 95% CI, 2.42-5.20) at previous sampling or sharing a towel with a MRSA-colonized individual (OR, 1.48; 95% CI, 1.14-1.92) were at higher risk for interval SSTI, as were those colonized with MRSA at more anatomic sites during previous sampling (relative risk, 2.33; $P < .001$) (Figure 2 and eFigure 3 in the [Supplement](#)).

Frequent cleaning of refrigerator handles was associated with fewer interval SSTIs (0.05 vs 0.09 SSTIs per person-sampling; $P = .03$) (eTable 13 in the [Supplement](#)). Households with at least 1 household member reporting an SSTI in the year before enrollment (not including the index patient's SSTI at enrollment) had more interval SSTIs (0.10 vs 0 SSTIs per person-sampling; $P < .001$). Higher personal MRSA colonization pressure (ρ , 0.21; $P < .001$) and environmental MRSA contamination pressure (ρ , 0.13; $P = .006$) were associated with more interval SSTIs across household members.

In the multivariable interval SSTI model, individuals persistently colonized with MRSA (OR, 1.56; 95% CrI, 1.17-2.11), those with an SSTI in the year before enrollment (excluding index enrollment infection; OR, 2.55; 95% CrI, 1.88-3.47), and index patients (OR, 1.54; 95% CrI, 1.07-2.23) were more likely

Figure 3. Markov Model of Strain Persistence Among Children, Adults, and the Environment



A 5-state Markov model was fit across all households and persistent strain types, with the resultant transition probabilities (and 95% CI estimates) re-scaled for the displayed 4-state model. Strains most commonly persist within households by persistent colonization of 1 individual or through transitions between adults or between children and less often through environment-only

persistence. Smaller people indicate at least 1 child (aged <18 years) colonized; larger people, at least 1 adult (aged ≥18 years) colonized; house, at least 1 environmental site colonized (but no household members colonized); and empty circle, strain type not present in household.

to report an interval SSTI (Table 2). Increasing environmental MRSA contamination pressure at the previous sampling (OR, 1.09; 95% CrI, 0.99-1.21) and sharing a bedroom with a MRSA-colonized individual (OR, 1.23; 95% CrI, 0.94-1.60) remained in the final model despite not reaching statistical significance (Figure 2A and eFigure 3 in the Supplement).

Discussion

The commensal state of *S aureus* challenges prevention efforts because colonization serves as a reservoir for endogenous infection, further perpetuating persistent colonization and transmission. This poses a significant burden, particularly in households.³¹ Although topical antimicrobials and antiseptics used by patients with SSTI have demonstrated eradication of *S aureus* carriage in decolonization trials, their effectiveness in preventing recurrent SSTI wanes over time, with more than 50% experiencing recurrence during 1 year.^{9,32-34} To develop more effective prevention strategies, it is essential to illuminate individual and household factors associated with persistent *S aureus* colonization and SSTI. The present work attempted to fill this knowledge gap by using a combination of longitudinal sampling of household members and their environments; high-resolution molecular typing of all recovered strains; detailed health, hygiene, and house-

hold behaviors; and tracking of incident SSTIs. We found that the index patient's infecting MRSA strain was most likely to persist in the household; that environmental contamination, but not pets, was associated with personal strain persistence; and that personal MRSA strain persistence was associated with increased likelihood of development of subsequent SSTI.

A study in Pennsylvania⁸ found that index patients with community-associated MRSA who were colonized with MRSA at SSTI onset remained colonized for a median of 140 days; colonization never cleared in 40% during the 6-month study period. Persons in whom colonization cleared were more likely to experience reacquisition if there was a higher proportion of children in the home or if they lived with a MRSA-colonized household contact.³⁵ Our study extended this knowledge because high environmental contamination burden (unmeasured in the previous study) was associated with individual persistent strain colonization. Moreover, although MRSA frequently colonized extranasal sites, nasal colonization was most likely to persist. Of interest, frequent handwashing alone was not associated with reduced persistence. Interval application of intranasal mupirocin was associated with protection against persistence, suggesting that measures beyond hand hygiene are necessary to interrupt persistent colonization. We found that the duration of persistent MRSA colonization was correlated with increased risk of SSTI in the modeled individual and in all household contacts.

Our study observed a difference in interval SSTI risk between individuals and households experiencing MRSA vs MSSA persistence. In households affected by community-associated MRSA SSTI, MRSA persistence was associated with increased likelihood of interval SSTI and MSSA persistence was inversely associated with SSTI incidence. Thus, strain competition, whether from noninfecting MRSA or MSSA strains, appears to oppose persistence of the MRSA infecting strain. Although contemporary community-associated SSTIs are most often attributable to MRSA, MSSA SSTIs are increasing.³⁶ In households of patients with MSSA SSTI, persistence dynamics and risk factors for SSTI may be different.

The present study reinforces emerging observations of the household as the primary source of community-associated MRSA, as suggested by an agent-based simulation of community-associated MRSA colonization in Chicago, Illinois, by Macal et al.³⁷ In our real-world study, we found no activities external to the household (secondary covariates in the multivariate models such as nail salons, sports, and occupation, as seen in eTables 2 and 3 in the [Supplement](#)) associated with persistent colonization or interval SSTI. We replicated the finding that individuals with an SSTI are at increased likelihood to experience subsequent SSTIs and cohabitate with others with a history of SSTI.^{9,17,38} The infecting strain at enrollment was the strain most likely to persist within a household, and its persistence was significantly associated with interval SSTI. In addition, strains causing interval SSTIs in household members were typically present within the household at prior sampling and were frequently concordant with the infecting strain at enrollment. Similarly, in a study conducted in Los Angeles, California, and Chicago, the infecting strain at enrollment was prevalent in approximately 50% of contaminated households for up to 3 months.¹⁹ These findings signify a cycle of MRSA colonization and infection in households: persistent personal MRSA colonization may lead to SSTI, and SSTI in turn may be associated with increased likelihood of MRSA acquisition and persistent colonization in household contacts.

We found that environmental MRSA contamination was associated with both strain-specific, personal-persistent colonization and interval SSTI. This finding is in contrast to the simulation by Macal et al³⁷ that posited that community-associated MRSA was spread solely through interpersonal contact. In addition, although previous studies in New York, NY,¹³ and in Los Angeles and Chicago¹⁷ found household fomite *S aureus* contamination to be independently associated with interval SSTIs in the index patient, the longitudinal sampling and

hierarchical modeling of our study assessed the associations among environmental contamination, persistent colonization, and interval SSTI. In our study, persistent colonization was reduced in households with higher cleanliness scores and frequent laundering of bathroom towels; cleaning of select surfaces was associated with reduced SSTI incidence. Taken together, our analyses suggest that long-term eradication and SSTI prevention measures should be targeted at both the personal and the household levels, pairing personal decolonization with environmental hygiene interventions associated with reduced contamination.

Strengths and Limitations

The strength of our study comes from its comprehensive modeling of clinical and molecular epidemiologic data in households (with 96% household member participation plus inclusion of pets) during a 12-month sampling time to provide a novel understanding of the association between persistent colonization and SSTI (with strain-type resolution), drawing from a diverse community sample. Previous studies lacked the temporal resolution, molecular resolution, environmental sampling, and/or sample size to provide a rigorous, longitudinal perspective of colonization persistence patterns and recurrent SSTI.^{8,13,17,19,35,39}

Limitations include that interval SSTI isolates were usually unavailable for molecular typing. In addition, our findings may not be generalizable to geographic locales with less prevalent MRSA colonization and infection or to households with SSTI caused by MSSA strains.

Conclusions

Recurrent SSTI is associated with strain-specific, persistent MRSA colonization of household members and contamination of environmental surfaces. Future randomized clinical trials are necessary to define the effectiveness of specific combinations of personal decolonization and environmental decontamination efforts in preventing recurrent SSTI in households. Our findings also suggest that the relative prevalence of other strains in the household may be associated with destabilization of a given strain's capacity to persist longitudinally on an individual. These data highlight the need for future studies of *S aureus* ecologic features and how other components of the skin and nasal microbiota may prevent or facilitate the transition of MRSA from colonization to infection.

ARTICLE INFORMATION

Accepted for Publication: January 8, 2020.

Published Online: March 30, 2020.

doi:10.1001/jamapediatrics.2020.0132

Author Affiliations: Department of Pediatrics, Washington University School of Medicine in St Louis, St Louis, Missouri (Hogan, Thompson, Muenks, Boyle, Sullivan, Morelli, Williams, Sanchez, Hunstad, Wardenburg, Burnham, Fritz); Graduate Program in the Biophysical Sciences, University of Chicago, Chicago, Illinois (Mork); Committee of Microbiology, University of Chicago, Chicago, Illinois

(Mork); Institute for Genomics and Systems Biology, University of Chicago, Chicago, Illinois (Mork); Department of Molecular Microbiology, Washington University School of Medicine in St Louis, St Louis, Missouri (Hunstad, Burnham); Department of Surgery, Washington University School of Medicine in St Louis, St Louis, Missouri (Gehlert); Department of Pathology and Immunology, Washington University School of Medicine in St Louis, St Louis, Missouri (Burnham); Department of Medicine, Washington University School of Medicine in St Louis, St Louis, Missouri

(Burnham); Department of Human Genetics, University of Chicago, Chicago, Illinois (Rzhetsky).

Author Contributions: Dr Fritz had full access to all of the data in the study and takes responsibility for the integrity of the data and the accuracy of the data analysis.

Concept and design: Hogan, Morelli, Hunstad, Bubeck Wardenburg, Gehlert, Burnham, Fritz.

Acquisition, analysis, or interpretation of data: Hogan, Mork, Thompson, Muenks, Boyle, Sullivan, Morelli, Williams, Sanchez, Hunstad, Gehlert, Burnham, Rzhetsky, Fritz.

Drafting of the manuscript: Hogan, Mork, Williams, Sanchez, Bubeck Wardenburg, Gehlert, Fritz.
Critical revision of the manuscript for important intellectual content: Hogan, Mork, Thompson, Muenks, Boyle, Sullivan, Morelli, Sanchez, Hunstad, Burnham, Rzhetsky, Fritz.

Statistical analysis: Hogan, Mork, Williams,

Sanchez, Gehlert, Rzhetsky, Fritz.

Obtained funding: Rzhetsky, Fritz.

Administrative, technical, or material support:

Hogan, Muenks, Sullivan, Morelli, Hunstad, Burnham, Fritz.

Supervision: Hogan, Burnham, Rzhetsky, Fritz.

Conflict of Interest Disclosures: Dr Hunstad reported receiving personal fees from BioVersys AG outside the submitted work. Dr Bubeck Wardenburg reported having a financial agreement with Ardis Pharmaceuticals related to patents owned by the University of Chicago. Dr Burnham reported receiving grants from Cepheid and bioMerieux and receiving personal fees from Thermo Fisher Scientific and Bio Rad Laboratories outside the submitted work. No other disclosures were reported.

Funding/Support: This work was supported by the Children's Discovery Institute of Washington University and St Louis Children's Hospital; grant K23-AI091690 (Dr Fritz) from the NIH/National Institute of Allergy and Infectious Diseases; grant UL1-TR002345 (Dr Fritz) from the NIH/National Center for Advancing Translational Sciences; grants R01-HS021736 and R01-HS024269 (Dr Fritz) from the Agency for Healthcare Research and Quality; and the Burroughs Wellcome Foundation Investigators in the Pathogenesis of Infectious Disease Award (Dr Bubeck Wardenburg). The computational analysis was partially funded by ARO contract W911NF1410333 (Dr Rzhetsky) from the Defense Advanced Research Projects Agency Big Mechanism program; grants R01HL122712, 1P50MH094267, and U01HL108634 (Dr Rzhetsky) from the NIH; and a gift from Liz and Kent Dauten (Dr Rzhetsky).

Role of the Funder/Sponsor: The funding agencies had no role in the design and conduct of the study; collection, management, analysis, and interpretation of the data; preparation, review, or approval of the manuscript; and decision to submit the manuscript for publication.

Disclaimer: The content is solely the responsibility of the authors and does not necessarily represent the official views of the NIH or the Agency for Healthcare Research and Quality.

Additional Contributions: Artwork used in Figure 2 was created by the following artists from the Noun Project (<https://thenounproject.com>): Xinh Studio (anterior nares); Gregory Montigny (axillae); Bohdan Burmich (inguinal folds); Oksana Latysheva (dog nose and dorsal fur); Priyanka (television remote control); Thays Malcher (main telephone); Patrick Morrison (computer keyboard and mouse); Andrea Severgnini (videogame controller); parkjusun (index patient bed linens); AomAm (hand towel); Ben Davis (sink faucet handle); Ralf Schmitzer (toilet seat and handle); Sara Jeffries (light switch); Halil Ibrahim Nuroglu (sink); Cecile Parker (bathtub); Llisole (countertop); Bakunetsu Kaito (soap bar and dish in bath or shower); Gabriele Fumero (door handle); icon 54 (index patient bath towel); Oleksandr Panasovskyi (sponge or cloth); dajeong lee (refrigerator door handle); Alexander Mravcak (table); Prettycons

(house); and Becris (cityscape). The following assisted with patient recruitment: Rachel Orscheln, MD, Washington University; Jennifer Seigel, RN, PNP, the St Louis Children's Hospital Pediatric Ambulatory Wound Service; Mary Bixby, RN, BSN, Cardinal Glennon Children's Hospital; Jane Garbutt, MB, ChB, Sherry Dodd, BA; and the physicians and staff of the participating Washington University Pediatric and Adolescent Ambulatory Research Consortium practices, including Mercy Clinic Pediatrics—Union, Missouri, and Washington Missouri; Johnson Pediatric Center, St Louis, Missouri; Heartland Pediatrics, Collinsville, Illinois; Forest Park Pediatrics, St Louis, Missouri; Tots Thru Teens, Bridgeton, Missouri; Pediatric Healthcare Unlimited, Alton, Illinois; Northwest Pediatrics—St Charles, St Charles, Missouri; Esse Health Pediatric & Adolescent Medicine—Watson Road, St Louis, Missouri; Fenton Pediatrics, LLC, Fenton, Missouri; Blue Fish Pediatrics, St Louis, Missouri; and Southwest Pediatrics, St Louis, Missouri. Meghan Wallace, BS, and Angela Shupe, BS, (both from Washington University School of Medicine in St Louis, St Louis, Missouri), provided technical assistance with molecular typing of *S aureus* isolates. Mike Talcott, DVM, and Mary Ellenberger, DVM, MS, (both from Washington University School of Medicine in St Louis, St Louis, Missouri) provided training in animal specimen collection. Susan Stark, PhD (Washington University School of Medicine in St Louis, St Louis, Missouri), provided guidance in conducting home study visits. All contributors received no financial compensation.

REFERENCES

- Kaplan SL. Community-acquired methicillin-resistant *Staphylococcus aureus* infections in children. *Semin Pediatr Infect Dis*. 2006;17(3):113-119. doi:10.1053/j.spid.2006.06.004
- Miller LG, Diep BA. Clinical practice: colonization, fomites, and virulence: rethinking the pathogenesis of community-associated methicillin-resistant *Staphylococcus aureus* infection. *Clin Infect Dis*. 2008;46(5):752-760. doi:10.1086/526773
- Dukic VM, Lauderdale DS, Wilder J, Daum RS, David MZ. Epidemics of community-associated methicillin-resistant *Staphylococcus aureus* in the United States: a meta-analysis. *PLoS One*. 2013;8(1):e52722. doi:10.1371/journal.pone.0052722
- Otto M. Community-associated MRSA: a dangerous epidemic. *Future Microbiol*. 2007;2(5):457-459. doi:10.2217/17460913.2.5.457
- Nouwen JL, Ott A, Kluytmans-Vandenbergh MF, et al. Predicting the *Staphylococcus aureus* nasal carrier state: derivation and validation of a "culture rule". *Clin Infect Dis*. 2004;39(6):806-811. doi:10.1086/423376
- Sollid JUE, Furberg AS, Hanssen AM, Johannessen M. *Staphylococcus aureus*: determinants of human carriage. *Infect Genet Evol*. 2014;21:531-541. doi:10.1016/j.meegid.2013.03.020
- Fritz SA, Epplin EK, Garbutt J, Storch GA. Skin infection in children colonized with community-associated methicillin-resistant *Staphylococcus aureus*. *J Infect*. 2009;59(6):394-401. doi:10.1016/j.jinf.2009.09.001
- Cluzet VC, Gerber JS, Nachamkin I, et al. Duration of colonization and determinants of earlier clearance of colonization with methicillin-resistant *Staphylococcus aureus*. *Clin Infect Dis*. 2015;60(10):1489-1496. doi:10.1093/cid/civ075
- Fritz SA, Hogan PG, Hayek G, et al. Household versus individual approaches to eradication of community-associated *Staphylococcus aureus* in children: a randomized trial. *Clin Infect Dis*. 2012;54(6):743-751. doi:10.1093/cid/cir919
- Fritz SA, Hogan PG, Hayek G, et al. *Staphylococcus aureus* colonization in children with community-associated *Staphylococcus aureus* skin infections and their household contacts. *Arch Pediatr Adolesc Med*. 2012;166(6):551-557. doi:10.1001/archpediatrics.2011.900
- Rodriguez M, Hogan PG, Krauss M, Warren DK, Fritz SA. Measurement and impact of *Staphylococcus aureus* colonization pressure in households. *J Pediatric Infect Dis Soc*. 2013;2(2):147-154. doi:10.1093/jpids/pit002
- Fritz SA, Hogan PG, Singh LN, et al. Contamination of environmental surfaces with *Staphylococcus aureus* in households with children infected with methicillin-resistant *S aureus*. *JAMA Pediatr*. 2014;168(11):1030-1038. doi:10.1001/jamapediatrics.2014.1218
- Knox J, Sullivan SB, Urena J, et al. Association of environmental contamination in the home with the risk for recurrent community-associated, methicillin-resistant *Staphylococcus aureus* infection. *JAMA Intern Med*. 2016;176(6):807-815. doi:10.1001/jamainternmed.2016.1500
- Ng W, Faheem A, McGeer A, et al. Community- and healthcare-associated methicillin-resistant *Staphylococcus aureus* strains: an investigation into household transmission, risk factors, and environmental contamination. *Infect Control Hosp Epidemiol*. 2017;38(1):61-67. doi:10.1017/ice.2016.245
- Hogan PG, Mork RL, Boyle MG, et al. Interplay of personal, pet, and environmental colonization in households affected by community-associated methicillin-resistant *Staphylococcus aureus*. *J Infect*. 2019;78(3):200-207. doi:10.1016/j.jinf.2018.11.006
- Mork RL, Hogan PG, Muenks CE, et al. Comprehensive modeling reveals proximity, seasonality, and hygiene practices as key determinants of MRSA colonization in exposed households. *Pediatr Res*. 2018;84(5):668-676. doi:10.1038/s41390-018-0113-x
- Miller LG, Eells SJ, David MZ, et al. *Staphylococcus aureus* skin infection recurrences among household members: an examination of host, behavioral, and pathogen-level predictors. *Clin Infect Dis*. 2015;60(5):753-763. doi:10.1093/cid/ciu943
- Nerby JM, Gorwitz R, Leshner L, et al. Risk factors for household transmission of community-associated methicillin-resistant *Staphylococcus aureus*. *Pediatr Infect Dis J*. 2011;30(11):927-932. doi:10.1097/INF.0b013e31822256c3
- Eells SJ, David MZ, Taylor A, et al. Persistent environmental contamination with USA300 methicillin-resistant *Staphylococcus aureus* and other pathogenic strain types in households with *S aureus* skin infections. *Infect Control Hosp Epidemiol*. 2014;35(11):1373-1382. doi:10.1086/678414
- Davis M, Morris D, Cluzet V, et al. Home environmental contamination is associated with community-associated methicillin-resistant *Staphylococcus aureus* re-colonization in treated patients. *Open Forum Infect Dis*. 2017;4(suppl 1):S7. doi:10.1093/ofid/ofx162.016

21. Mork RL, Hogan PG, Muenks CE, et al. Longitudinal, strain-specific *Staphylococcus aureus* introduction and transmission events in households of children with community-associated methicillin-resistant *S aureus* skin and soft tissue infection: a prospective cohort study. *Lancet Infect Dis*. 2020;20(2):188-198. doi:10.1016/S1473-3099(19)30570-5
22. Halliday G, Snowdon J. The Environmental Cleanliness and Clutter Scale (ECCS). *Int Psychogeriatr*. 2009;21(6):1041-1050. doi:10.1017/S1041610209990135
23. Hogan PG, Burnham CA, Singh LN, et al. Evaluation of environmental sampling methods for detection of *Staphylococcus aureus* on fomites. *Ann Public Health Res*. 2015;2(1):1013.
24. Cockerill F. *Performance Standards for Antimicrobial Susceptibility Testing: Twenty-third Informational Supplement*. Clinical and Laboratory Standards Institute; 2013:M100-S23.
25. Rodriguez M, Hogan PG, Satola SW, et al. Discriminatory indices of typing methods for epidemiologic analysis of contemporary *Staphylococcus aureus* strains. *Medicine (Baltimore)*. 2015;94(37):e1534. doi:10.1097/MD.0000000000001534
26. Del Vecchio VG, Petroziello JM, Gress MJ, et al. Molecular genotyping of methicillin-resistant *Staphylococcus aureus* via fluorophore-enhanced repetitive-sequence PCR. *J Clin Microbiol*. 1995;33(8):2141-2144. doi:10.1128/JCM.33.8.2141-2144.1995
27. Virtanen P, Gommers R, Oliphant TE, et al; SciPy 1.0 Contributors. SciPy 1.0: fundamental algorithms for scientific computing in Python. *Nat Methods*. Published online February 3, 2020. doi:10.1038/s41592-019-0686-2
28. R Foundation for Statistical Computing. R: a language and environment statistical computing. Published 2016. Accessed September 18, 2018. <https://www.R-project.org/>
29. Hadfield J. MCMC methods for multi-response generalized linear mixed models: the MCMCglmm R package. *J Stat Softw*. 2010;33(2):1-22. doi:10.18637/jss.v033.i02
30. Jackson C. Multi-state models for panel data: the msm package for R. *J Stat Softw*. 2011;38(8):1-28. doi:10.18637/jss.v038.i08
31. Chisholm RH, Campbell PT, Wu Y, Tong SYC, McVernon J, Geard N. Implications of asymptomatic carriers for infectious disease transmission and control. *R Soc Open Sci*. 2018;5(2):172341. doi:10.1098/rsos.172341
32. Fritz SA, Camins BC, Eisenstein KA, et al. Effectiveness of measures to eradicate *Staphylococcus aureus* carriage in patients with community-associated skin and soft-tissue infections: a randomized trial. *Infect Control Hosp Epidemiol*. 2011;32(9):872-880. doi:10.1086/661285
33. Ellis MW, Griffith ME, Dooley DP, et al. Targeted intranasal mupirocin to prevent colonization and infection by community-associated methicillin-resistant *Staphylococcus aureus* strains in soldiers: a cluster randomized controlled trial. *Antimicrob Agents Chemother*. 2007;51(10):3591-3598. doi:10.1128/AAC.01086-06
34. Kaplan SL, Forbes A, Hammerman WA, et al. Randomized trial of "bleach baths" plus routine hygienic measures vs. routine hygienic measures alone for prevention of recurrent infections. *Clin Infect Dis*. 2014;58(5):679-682. doi:10.1093/cid/cit764
35. Cluzet VC, Gerber JS, Nachamkin I, et al. Risk factors for recurrent colonization with methicillin-resistant *Staphylococcus aureus* in community-dwelling adults and children. *Infect Control Hosp Epidemiol*. 2015;36(7):786-793. doi:10.1017/ice.2015.76
36. Sutter DE, Milburn E, Chukwuma U, Dzialowy N, Maranich AM, Hospenhal DR. Changing susceptibility of *Staphylococcus aureus* in a US pediatric population. *Pediatrics*. 2016;137(4):e20153099. doi:10.1542/peds.2015-3099
37. Macal CM, North MJ, Collier N, et al. Modeling the transmission of community-associated methicillin-resistant *Staphylococcus aureus*: a dynamic agent-based simulation. *J Transl Med*. 2014;12:124. doi:10.1186/1479-5876-12-124
38. Miller LG, Eells SJ, Taylor AR, et al. *Staphylococcus aureus* colonization among household contacts of patients with skin infections: risk factors, strain discordance, and complex ecology. *Clin Infect Dis*. 2012;54(11):1523-1535. doi:10.1093/cid/cis213
39. Rodriguez M, Hogan PG, Burnham C-A, Fritz SA. Molecular epidemiology of *Staphylococcus aureus* in households of children with community-associated *S aureus* skin and soft tissue infections. *J Pediatr*. 2014;164(1):105-111. doi:10.1016/j.jpeds.2013.08.072